KURUKSHETRA UNIVERSITYKURUKSHETRA

Syllabus forUnder-Graduate

Programme

Course: Bachelor of Vocation in Medical Laboratory Technology (3rd & 4th semester)

Under Multiple Entry-Exit, Internship and CBCS-LOCF in accordance to NEP-2020 w.e.f. 2023-24

PLOs for Life Sciences

PLOs	UG Certificate in Life Sciences
After the comple	etion of UG certificate in Life Sciences, the student should be able to:
PLO_1: Knowledge and	• Demonstrate the knowledge of basic principles, concepts, facts and broad
Understanding	linkage of chosen subjects of Life sciences.
PLO_2: Skills	• Selecting and using relevant methods, tools, and materials to assess the
And creativity	appropriateness of approaches for solving problems associated with the
	chosen subjects of Life sciences.
PLO_3: Application of	• Apply the acquired operational or theoretical knowledge, and a range of
knowledge and Skills	practical skills to select and use basic methods, tools, materials, and
	information to generate solutions to specific problems relating to the chosen
	subjects of Life sciences.
PLO_4: Critical	• Listen carefully, read texts, make judgments and take decisions based on
thinking	analysis of data and evidences.
PLO_5: Ethics	• Put forward convincing arguments to respond to the ethical and moral issues
	associated with the chosen subjects, practice ethical and moral values in one's life.
PLO_6: Communication	• Express scientific thoughts and ideas effectively in writing and orally and communicate on scientific activities with others using appropriate media.
PLO_7:Life Long Learning	• Acquire knowledge and skills including learning 'How to learn' that are necessary for participating in learning activities throughout life.
PLO_8: Environmental Awareness	• Demonstrate knowledge of effects of environmental degradation, climate change and pollution, effective waste management.
PLO_9:Digital Literacy	• To use ICT in a variety of learning and work situations.

PLOs	UG Diploma in Life Sciences
After the comp	letion of UG Diploma in Life Sciences, the student should be able to:
PLO_1: Knowledge and Understanding	• Demonstrate the deeper knowledge and understanding of principles, concepts, facts and broad linkage of chosen subjects of Life sciences.
PLO_2: Skills And creativity	• Selecting and using relevant methods, tools, and materials to assess the appropriateness of approaches from a range of sources for solving complex problems associated with the chosen subjects of Life sciences.
PLO_3: Application of knowledge and Skills	• Apply the acquired operational or theoretical knowledge, and a range of practical skills to select and use appropriate methods, tools, materials, and information to generate solutions to specific problems relating to the chosen subjects of Life sciences.
PLO_4: Critical thinking	• Listen carefully, read texts, make judgments and take decisions based on analysis of data and evidences, present complex information in a clear, scientific and concise manner.
PLO_5: Ethics	• Formulate arguments in support of actions to address the ethical and moral issues associated with the chosen subjects, practice ethical and moral values in one's life.
PLO_6: Communication	• Express scientific thoughts and ideas effectively in writing and orally and communicate on scientific activities with others using appropriate media.
PLO_7:Life Long Learning	• Acquire knowledge and skills including learning 'How to learn' that are necessary for participating in learning activities throughout life.
PLO_8: Environmental Awareness	• Apply knowledge, skills and attitude to mitigate the effects of environmental degradation, climate change and pollution, effective waste management.
PLO_9:Digital Literacy	• To use ICT in a variety of learning and work situations.

PLOs	Bachelor Degree in Life Sciences
After the complet	tion of Bachelor degree in Life Sciences, the student should be able to:
PLO_1: Knowledge and Understanding	• Demonstrate the comprehensive and specialized knowledge and deep understanding of principles, concepts, and factsabout current and emerging issues relevant to chosen subjects of Life sciences.
PLO_2: Skills And creativity	• Selecting and using relevant methods, tools, and materials to assess the appropriateness of approaches for solving specific problems associated with the chosen subjects of Life sciences.
PLO_3: Application of knowledge and Skills	• Apply the acquired operational or theoretical knowledge, and a range of practical skills to analyze quantitative and qualitative data to assess the different approaches to generate solutions to specific problems related to the chosen subjects of Life sciences.
PLO_4: Critical thinking	• Listen carefully, read texts, make judgments and take decisions based on analysis of data and evidences, present complex information in a clear, scientific and concise manner.
PLO_5: Ethics	• Follow ethical practices in all aspects of research and development, including avoiding unethical practices such as fabrication, falsification or misrepresentation of data or committing plagiarism.
PLO_6: Communication	• Able to communicate effectively on complex scientific activities with the scientific community and with society at large, such as, being able to comprehend and write effective scientific reports and design documentation, make effective presentations.
PLO_7:Life Long Learning	• Acquire knowledge and skills including learning 'How to learn' that are necessary for participating in learning activities throughout life.
PLO_8: Environmental Awareness	• Apply knowledge, skills and attitude to mitigate the effects of environmental degradation, climate change and pollution, effective waste management.
PLO_9:Digital Literacy	• To use ICT in a variety of learning and work situations, appropriate software to analysis the data.
PLO_10:Research Aptitude	• Ask relevant/appropriate questions, identifying, formulating and analyzing the research problems and to draw conclusion from the analysis.

<u>CC-A3</u>

	Session:2024-25		
	Part A-Introduction		
Subject	Bachelor of Vocation in Med	lical Laboratory Techr	ology
Semester	III		
Name of the Course	Biochemistry–III		
Course Code	B23-MLT-301		
Course Type:	CC		
Level of the course (As per Annexure-I	200-299		
Pre-requisite for the course (if any)			
Course Learning Outcomes (CLO): CLO5 is based on practical component	 equipment. Introduce the students analytical and diagnos Understand the print important techniques Impart general insight various procedures lik Provide skills for according to the students of the students of	tical techniques by usi to various apparatus stic sections of Biocher nciple procedures an like ELISA. t into theoretical and p the PCR. urate laboratory results rd experimental con	ng sophisticated and reagents used in mistry. nd applications of ractical education of s in a timely manner
Credits	Theory	Practical	Total
	03	01	04
Contact Hours	03	02	05
Max.Marks:100 Internal Assessment Marks:30 End Term Exam Marks: 70 (Th	•	Exam duration: Th Practical:4 hours	neory:3 Hours
	Part B- Contents of the C	Course	
Instructions for Paper-Setter: Nine questions will be set in all. the entire syllabus, will be comp			

Nine questions will be set in all. Question No.1 comprising of objective/short answer type questions from the entire syllabus, will be compulsory. The remaining eight questions will be set taking two questions from each unit. The candidates will be required to attempt Q.No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact
		Hours
Ι	Analytical Biochemistry-I : Titrimetry, Colorimetry, Spectrophotometry, Flame	10
	photometry, Atomic absorption spectroscopy, Automatic washer.	
II	Analytical Biochemistry-II : Electrometric determination of Na ⁺ and K ⁺ ,	10
	Chromatography (Thin-layer and liquid chromatography), Electrophoresis	
	(Paper and Gel electrophoresis) for haemoglobin.	
III	Principle procedures and applications of :	12
	(a) Semi auto-analyzer, diluters and dry chemistry analyzer.	
	(b) Osmometry	
	(d) ELISA (Enzyme Linked Immunosorbent Assay)	
IV	Principle procedures and applications of :	13
	(a) Coulter-counters	
	(b) RIA (Radio-Immunoassay)	
	(c) PCR (Polymerase Chain Reaction)	
V*	PRACTICAL	30
	1. Demonstration of principle and working of: Colorimeter,	
	spectrophotometer, flame photometer, PCR (Polymerase Chain	
	Reaction), coulter-counters.	
	2. Demonstration of osmometry.	
	3. Demonstration of RIA(Radio-Immunoassay)	
	4. Demonstration of DOT ELISA (Enzyme Linked Immunosorbent Assay)	
	5. Demonstration of polyacrylamide Gel Electrophoresis of a biological sample.	

Suggested Evaluation Methods					
Internal Assessment: ≻ Theory	End Term Examination:				
 Class Participation: 5 Seminar/presentation/assignment/quiz/classtestetc.:5 Mid-Term Exam: 10 	Theory: 50 (Written exam) Practical: 20 (Seminar/				
 Practicum Class Participation: NA Seminar/Demonstration/Viva-voce/Labrecordsetc.:10 Mid-Term Exam: NA 	Demonstration/ Viva-voce/Lab records etc)				

	Part C- Learning Resources
Rec	commended Books/e-resources/LMS:
1.	Textbooks of Medical Laboratory Technology, Volume 1, 3rd Edition by Praful Ghodkar.
2.	Textbook of Medical Laboratory Technology, Volume 2, 3rd Edition by Praful Ghodkar.
2	Madical Laboratory Technology (Valume D), Dreadure Manual for Deutine Discussion Kanai

- Medical Laboratory Technology (Volume I) : Procedure Manual for Routine Diagnostic, Kanai Mukharjee.
- 4. Medical Laboratory Technology (Volume 2): Procedure Manual for Routine Diagnostic, Kanai Mukherjee.
- Medical Laboratory Technology (Volume 3): Procedure Manual for Routine Diagnostic, Kanai Mukherjee.

	PLOCLO Mapping of B23-MLT-301								
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9
CLO1	1	1	1	1	1	1	1	1.5	1
CLO2	1.5	1	2	1	1	1.5	1	1.5	1
CLO3	1	1	2	1.5	1.5	1.5	1	1.5	1
CLO4	1.5	1	2	1.5	1.5	1	1	1.5	1
CLO5	1	1	2	1.5	1.5	1	1	1.5	1

	CC-B3					
	Session:2024-25					
	Part A-Introduction					
Subject	Bachelor of Vocation in I	Medical Laboratory Tech	nology			
Semester	Ш	Ш				
Name of the Course	Microbiology-III					
Course Code	B23-MLT-302					
Course Type:	CC					
Level of the course (As per Annexure-I	200-299					
Pre-requisite for the course (if any)						
Course Learning Outcomes (CLO):	 After completing this course, the learner will be able to: 1. Demonstrate the knowledge of various micro-organisms. 2. Learn the general characters, diagnosis and life-cycles of medically important micro-organisms. 3. Provide the student methods with the study of staining techniques. 4. Gain knowledge about the etiological agents responsible for global infectious diseases. 					
CLO5isbasedonpractical component	5. Get an idea of univ	versal safety precautions.				
Credits	Theory	Practical	Total			
	03	01	04			
Contact Hours	03	02	05			
Max.Marks:100 Internal Assessment Marks: 30(Th End Term Exam Marks: 70 (Theor	•	Exam duration: The Practical:4 hours	ory:3 Hours			
	Part B-Contents of the C	Course				
Instructions for Donor Sotton						

Instructions for Paper-Setter:

Nine questions will be set in all. Question No.1 comprising of objective/short answer type questions from the entire syllabus, will be compulsory. There remaining eight questions will be set taking two questions from each unit. The candidates will be required to attempt Q. No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact Hours
Ι	Identification of Bacteria: Micrococci, Staphylococci, Streptococci, Pneumococci, Corynebacteria, Escherichia, Klebsiella, Enterobacter, Salmonella, Shigella, Citrobacter, Yersinia, Pseudomonas, Vibrio, Haemophilus, Mycoplasma, Rickettsia.	10
II	Pathogenic and Non-Pathogenic Fungi: Candida, Cryptococci, Dermatophytes, Sporothrix, Histoplasma, Blastomyces, Coccidioides, Para-coccidioides, Dematiaceous fungi, Mycetoma, Actinomyces, Nocardia.	10
Ш	Principles of staining techniques, composition & preparation of stains: Making of films, staining methods, mounting media. Gram stain - preparation of stain and staining methods, speical stains for acid fast bacilli (AFB), Diptheria, intracytoplamic lipids, polysaccharides, nuclear materials, stain for amoeba, fungi, rickettsia. Ziehl - Neelsen stain, Albert stain and negative stain.	12
IV	Morphology, life cycle and laboratory diagnosis of haemoflagellates (<i>Leishmania, Trypanosomes</i>); Morphology and life cycle of tissue and blood nematodes (<i>Trichinella</i> ; <i>Filaria; Dracunculus</i>), Lab. Diagnosis of tissue & blood nematode infection; Morphology and life cycle of intestinal cestodes (<i>Taenia, Echinococcus</i>); Culture techniques for protozoa (<i>Amoeba, Giardia, Leishmania</i>) ; Culture methods for Helminths, Hookworm, Round worm; Egg counting techniques; Preparation of stains and staining procedures of malaria; Identification of different plasmodium species; Preparation of media and maintenance of cultures (<i>E. histolytica; Giardia; Leishmania</i>)	13
V*	 PRACTICALS 1. Identification of Pathogenic and non-pathogenic fungi from class-work materials/diseased tissues. 2. Isolation of Lactobacilli and Streptococci from curd. 3. Morphology and Life cycle of haemoflagellates <i>Leishmania</i>, <i>Trypanosomes</i> 4. Laboratory diagnosis of <i>Leishmania</i>, <i>Trypanosomes</i>. 5. Morphology and life cycle of tissue and blood nematodes <i>Filaria</i>, <i>Trichinella</i>, <i>Dracunculus</i> 6. Lab. diagnosis of tissue & blood nematode infection. 7. Morphology and life cycle of intestinal cestodes <i>Taenia</i>, <i>Echinococcus</i> 8. Culture techniques for protozoa <i>Amoeba</i>, <i>Giardia</i>, <i>Leishmania</i> 9. Culture methods for Helminths, Hookworm, Round worm. 10. Preparation of stains and staining procedures of malaria. 11. Identification of different plasmodium species. 12. Preparation of media and maintenance of cultures of <i>E. histolytica</i>, <i>Giardia</i>, <i>Leishmania</i> 	30

Suggested Evaluation Methods	
Internal Assessment: ➤ Theory • Class Participation:5 • Seminar/presentation/assignment/quiz/classtestetc.:5 • Mid-Term Exam: 10 ➤ Practicum • Class Participation: NA • Seminar/Demonstration/Viva-voce/Labrecordsetc.:10 • Mid-Term Exam: NA	End Term Examination: Theory: 50 (Writter exam) Practical: 20 (Seminar/ Demonstration/Viva- voce/Lab records etc)
Part C-Learning Resources	
 Recommended Books/e-resources/LMS: 1. Practical and Applied Microbiology Anuradha De 4th Edition 2. Text Book of Microbiology Anantnarayan 10th Edition 3. Text Book of Microbiology and Parasitology Praful Godkar 1st Edition 4. Medical Parasitology C.P. Baweja 3rd Edition 5. Text Book of Microbiology for Nursing Students, Anant Narayan Panikar 6. Text Book of Ophthalmology, Khurana 7. Text Book of Microbiology, Baveja. 	

	PLOCLO Mapping of B23-MLT-302								
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9
CLO1	1	1	1	1	1	1.5	1	1.5	1
CLO2	1.5	1	2	1	1.5	1.5	1	1.5	1
CLO3	1	1	2	1.5	1	1.5	1	1.5	1.5
CLO4	1.5	1	1	1	1	1.5	1	1.5	1
CLO5	1	1	1	1	1	1.5	1	1.5	1.5

	Session:2024-25			
	Part A-Introduction	l		
Subject	Bachelor of Vocation	in Medical Laboratory	/ Technology	
Semester	Ш			
Name of the Course	Pathology–1II			
Course Code	B23-MLT-303			
Course Type:	CC			
Level of the course (As per Annexure-I	200-299			
Pre-requisite for the course (if any)				
Course Learning Outcomes (CLO):	 After completing this course, the learner will be able to: Gain the basic knowledge of cytology and Cytopathology. Impart sound knowledge and basic skills of working in a pathology lab. Understand the principle, procedure and demonstration of various tissue constituents and advanced tools. Diagnose routine and complex clinical problems on the basis of cytopathology specimens. 			
CLO5isbasedonpracticalcomponent	5. Enable the stud under guidance	dent to collect blood an e.	d urine samples	
Credits	Theory	Practical	Total	
	03	01	04	
Contact Hours	03	02	05	
Max.Marks:100 Internal Assessment Marks:30 (Theor End Term Exam Marks: 70 (Theory 5		Exam duration: Th Practical:4 hours	eory:3 Hours	
Pa	art B-Contents of the C	Course		

CC-C3

Instructions for Paper-Setter:

Nine questions will be set in all. Question No.1comprising of objective/short answer type questions from the entire syllabus, will be compulsory. The remaining eight questions will be set taking two questions from each unit. The candidates will be required to attempt Q.No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact Hours
I	General properties of living organisms; chemistry of the cells; cellular membranes; cytoskeleton; endoplasmic reticulum; golgi body; lysosomes; nuclear envelope; chromatin and chromosomes; mitosis; meiosis; Gametogenesis; reproductive cycle; fertilisation; cleavage; a model of gastrulation; Epithelial tissue; connective tissues (blood connective, cartilage, bone); muscular tissue; nervous tissue.	10
II	Introduction, evaluation and reporting of cytopathology specimens; Clinical residents in the following, keeping in view the special requirements of each case (Cytohormonal status, malignancy, infection, etc.); Types of smears (morning specimen, evening specimen, pre-menstrual specimen, etc.).	10
III	Method of obtaining various specimens : urine sample, gastric smear, colonic lavage etc. Principles and preparation of solutions of stains. Techniques for concentration of specimens : various filters and cytocentrifuge.	12
IV	Normal anatomy, histology and cytology of the cervix and endometrium, Sampling methods for the cervix, Microbiology of the female genital tract and the cytological preparations of common infections of the cervix. Introduction to routine screening and reporting of non-gynaecological cytology specimens including those from : Respiratory system, Urinary system.	13
V*	 PRACTICALS To study collection of specimens and their clinical significance. Preparation of specimens for cytological evaluation. Concentrating specimens by centrifugation Thick specimen Watery specimen To study preparation of smear, procedure of its fixation and mailing of smears To study cell division from prepared slides of mitosis and meiosis. Temporary squash preparations of onion root tip for the study of mitosis using acetocarmine stain. Gynaecological and non-gynaecological cytology preparations and their studies. 	30

Suggested Evaluation Methods					
Internal Assessment:	End Term				
≻ Theory	Examination:				
Class Participation: 5	Theory:50				
	(Written exam)				
 Seminar/presentation/assignment/quiz/classtestetc.:5 	Practical: 20 (Seminar/				
• Mid-Term Exam: 10	Demonstration/Viva-				
> Practicum	voce/Lab records etc)				
Class Participation: NA					
• Seminar/Demonstration/Viva-voce/Labrecordsetc.:10					
• Mid-Term Exam: NA					

Recommended Books/e-resources/LMS:

- 1. A Handbook of Medical Technology-second edition, BY V.H. Talib, CBS Publishers
- 2. Textbook of Medical Laboratory Technology, Volume 1, 3rd Edition by Praful Ghodkar
- 3. Textbook of Medical Laboratory Technology, Volume 2, 3rd Edition by Praful Ghodkar
- 4. Medical Laboratory Technology (Volume 1): Procedure Manual for Routine Diagnostic, Kanai Mukharjee
- 5. Medical Laboratory Technology (Volume 2): Procedure Manual for Routine Diagnostic, Kanai Mukharjee
- 6. Medical Laboratory Technology (Volume 3): Procedure Manual for Routine Diagnostic, Kanai Mukharjee

	PLOCLO Mapping of B23-MLT-303								
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9
CLO1	1	1.5	1	1	1	1.5	1	1.5	1.5
CLO2	1.5	1	2	1	1	1.5	1	1.5	1
CLO3	1	1.5	2	1	1	1.5	1	1.5	1.5
CLO4	1.5	1	1	1	1	1.5	1	1.5	1
CLO5	1	1.5	1	1	1	1.5	1	1.5	1.5

	Session:2024-25				
	Part A–Introduction	l			
Subject	Bachelor of Vocation in	Medical Laboratory T	echnology		
Semester	IV				
Name of the Course	Biochemistry–IV				
Course Code	B23-MLT-401				
Course Type:	CC				
Level of the course (As per Annexure-I	200-299				
Pre-requisite for the course (if any)					
Course Learning Outcomes (CLO): CLO5 is based on practical componen	 medical laboratory teo Able to do routine and laboratory practice. Teach about various r of various fluids and b Introduce the conce experimental control records. Understand the skill operation of lab in 	oretical and practical shall help students to chnology and science. d special investigative netabolic processes in pasic concept of nutriti pt of internal qualit s and management	education in clinical work in the field of procedures in medical the body, composition on. y control, safeguard and maintenance of s diagnostic profiles		
Credits	laboratory results. Theory	Practical	Total		
	03	01	04		
Contact Hours	03	02	05		
Max.Marks:100 Internal Assessment Marks: 30 (Th End Term Exam Marks: 70 (Theory	•	Exam duration: Tl Practical:4 hours	neory:3 Hours		
	Part B- Contents of the (Course			
Instructions for Paper-Setter: Nine questions will be set in all. Qu the entire syllabus, will be compulse from each unit. The candidates will	ory. The remaining eight	questions will be set	taking two questions		

CC-A4

from each unit. The candidates will be required to attempt Q.No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact Hours
I	Carbohydrae metabolism, glycolysis, Kreb cycle (Tricarboxylic acid cycle) and their clinical importance, glucose tolerance test (GTT). Protein metabolism-urea cycle and its biomedical significance, Lipid metabolism, Beta-oxidation of fatty acids, ketonebodies, metabolic changes in liver and adipose tissues during starvation, lipid profile.	10
Π	 Principle, assay procedures and clinical significance of following; Glucose, total proteins, A/G ratio, Albumin, globulin, urea, Blood Urea Nitrogen level (BUN), uric acid, creatinin, cholesterol, Billirubin (Direct and Indirect). Essential Electrolytes : Quantitative estimation of Sodium, potassium, calcium, chloride, lithium, phosphorus, magnesium inorganic phosphate, Protein Bound iodine (PBI) 17 Ketosteroids, Barbiturates and their clinical significance. 	12
III	Principle techniques and clinical significance of acid base balance test, D- Xylose absorption test, Inulin clearance test, urea and creatinin clearance tests, renal function tests, glycosylated haemoglobin & Liver function tests.	10
IV	Collection and preservation of biological fluids (blood, serum, plasma, urine and cerebrospinal fluid (CSF), Quality control of clinical investigation, normal ranges of various bio-metabolites and their confidence limits, automation in clinical biochemistry laboratory, laboratory organization management and maintenance of records.	13
V*	 PRACTICALS 1. Separation of sugars by Paper Chromatography. 2. Separation of lipids by thin-layer chromatography. 3. Determination of total soluble sugars by ferricyanide method (Colorimetric method). 4. Quantitative estimation of glucose in blood plasma using glucose oxidase by enzymatic method. 5. Determination of glycogen in liver. 6. Estimation of protein by Lowry's method. 7. Determination of sodium and potassium content in blood serum samples by flame photometer. 9. Quantitative estimation of calcium by volumetric procedure. 10. Quantitative analysis of lipids : (a) The determination of the acid value of a fat. (b) The saponification value of a fat. (c) The iodine number of a fat. (d) The estimation of blood cholesterol. 11. Estimation of blood sugar, urea, uric acid, creatinin, bilirubin etc. 	30

Suggested Evaluation Methods				
Internal Assessment: ➤ Theory • Class Participation: 5 • Seminar/presentation/assignment/quiz/classtestetc.:5 • Mid-Term Exam: 10 ➤ Practicum • Class Participation: NA • Seminar/Demonstration/Viva-voce/Labrecordsetc.:10 • Mid-Term Exam: NA	End Term Examination: Theory: 50 (Written exam) Practical: 20 (Seminar/ Demonstration/ Viva- voce/Lab records etc)			

Recommended Books/e-resources/LMS:

- 1. Essentials of Biochemistry, Second Edition, Dr.(Prof.) Satyanarayan
- 2. Essentials of Biochemistry, 2nd Edition, Dr. Pankaja Naik
- 3. Principles and Techniques of Biochemistry and Molecular Biology, 5th Edition, Wilson & Walker
- 4. An Introduction to Chemistry, 8th Edition by Mark Bishop
- 5. Clinical Chemistry made easy, 1st Edition by Hughes
- 6. Tietz Fundamentals of Clinical Chemistry, 7th Edition by Carl Burtis

	PLOCLO Mapping of B23-MLT-401								
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9
CLO1	1	1.5	1	1	1	1.5	1	1.5	1.5
CLO2	1.5	1	2	1	1	1.5	1	1.5	1
CLO3	1	1.5	2	1	1	1.5	1	1.5	1.5
CLO4	1.5	1	1	1	1	1.5	1	1.5	1
CLO5	1	1.5	1	1	1	1.5	1	1.5	1.5

	Session:2024-25				
	Part A-Introduction				
Subject	Bachelor of Vocation in	n Medical Laboratory	Fechnology		
Semester	IV				
Name of the Course	Microbiology-IV				
Course Code	B23-MLT-402				
Course Type:	CC				
Level of the course (As per Annexure-I	200-299				
Pre-requisite for the course (if any)					
Course Learning Outcomes (CLO): CLO5isbasedonpracticalcomponent	 After completing this course, the learner will be able to: Impart theoretical as well as practical training in various branches of microbiology namely virology, parasitology and bacteriology. Knowledge about the equipment used in microbiology, universal safety precautions and quality control measures. Gain directions for the care of animals, common diseases and experimental procedures to deal with animals. Know about the normal and pathogenic microorganisms, methods for recovery antibiotic testing and sterilization techniques. Gain knowledge of computers for the computation of laboratory results. 				
Credits	Theory	Practical	Total		
	03	01	04		
Contact Hours	03	02	05		
Max.Marks:100 Internal Assessment Marks:30(Theo End Term Exam Marks: 70 (Theory	•	Exam duration: Th Practical:4 hours	neory:3 Hours		

CC-B4

Part B-Contents of the Course

Instructions for Paper-Setter:

Nine questions will be set in all. Question No.1comprisingof objective/short answer type questions from the entire syllabus, will be compulsory. The remaining eight questions will be set taking two questions from each unit. The candidates will be required to attempt Q.No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact Hours			
Ι	Safety measure in Microbiology Laboratory : Occurence of La infections, route of infection in laboratory, safety measure precaution is use of pathogens in teaching. Lab organization managment, recording of results and quality control in Medical Microbiology Lab.	in			
Π	II Preservation of microbes and lyophilisation methods; Total viable count of microorganisms (bacteria, yeast & moulds); Testing of disinfectants : Rideal - Walker, Chick - Martin and In-use tests; Preparation and standardization of vaccines and immunization schedule; Sterilization - Definition, methods, principles, bacteriological filtration, irradiation, tyndalization.				
III	III Virology: Definition, General introduction of Virus, physico-chemical characteristics of viruses, diseases caused by different viruses and mode of infection, mode of transmission of viral agents, different staining techniques used in virology, use of embryonated eggs in clinical virology, principles of animal cell culture and their use in virology, use of common laboratory animals in viral diagnosis, prevention of viral diseases, immunity in viral infection.				
IV	IVCare and management of experimental animals: General directions for the care of animals, common diseases and experimental procedures. Various experimental animals - rabbits, guineapigs, mice, rats, fowls, and monkeys - their data, cages, feeding and handling.				
V*	V* PRACTICALS 1. Handling and care of laboratory animals. 2. Recording of laboratory data and use of computers. 3. Safety measures in Microbiology laboratory 4. Methods of preservation of microbes. 5. Isolation of bacteria from curd. 6. Indole, Methyl red, Voges-Proskauer, Citrate utilization (IMViC) test from coliform bacteria. 7. Study of sterilization techniques.				
	Suggested Evaluation Methods				
> Th • (• 2 • 1 > Pr • (• 2	Internal Assessment:Ex➤ TheoryEx• Class Participation: 5Theory• Seminar/presentation/assignment/quiz/classtestetc.:5Theory• Mid-Term Exam: 10Pho➤ PracticumD• Class Participation: NAVol• Seminar/Demonstration/Viva-voce/Labrecordsetc.:10Vol• Mid-Term Exam: NAVol				

Recommended Books/ e-resources/LMS:

- 1. Medical Parasitology C.P. Baweja 3rd Edition
- 2. Practical and Applied Microbiology Anuradha De 4th Edition 1
- 3. Text Book of Microbiology and Parasitology Praful Godkar 1st Edition
- 4. Parasitology in relation to Clinical Medicine by K.D. Chhatterjee
- 5. Concise Microbiology C.P. Baweja 1st Edition
- 6. Text Book of Microbiology Anantnarayan 10th Edition

PLOCLO Mapping of B23-MLT-402									
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9
CLO1	1	1.5	1	1	1	1.5	1	1.5	1.5
CLO2	1.5	1	2	1	1	1.5	1	1.5	1
CLO3	1	1.5	2	1	1	1.5	1	1.5	1.5
CLO4	1.5	1	1	1	1	1.5	1	1.5	1
CLO5	1	1.5	1	1	1	1.5	1	1.5	1.5

	Session:2024-25				
	Part A-Introduction	1			
Subject	Bachelor of Vocation	n in Medical Laboratory	y Technology		
Semester	IV				
Name of the Course	Pathology– IV				
Course Code	B23-MLT-403				
Course Type:	CC				
Level of the course (As per Annexure-I	200-299				
Pre-requisite for the course (if any)					
Course Learning Outcomes (CLO):	 Provide generation Gain knowled metachromasi Learn about cl Introduce the laboratory. Perform the 	s course, the learner will al insight into the basic dge about metal imp a and metachromatic dy assification of tissues a concept of quality techniques and state and cytology.	s of cytology. regnation techniques, yes. and their functions. control in cytology		
CLO5isbasedonpractical component					
Credits	Theory	Practical	Total		
	03	01	04		
Contact Hours	03	02	05		
Max.Marks:100 Internal Assessment Marks:30(Theory End Term Exam Marks: 70 (Theory 5		Exam duration: Th Practical:4 hours	neory:3 Hours		
Pa	art B-Contents of the	Course			
Instructions for Paper-Setter:					

CC-C4

Nine questions will be set in all. Question No.1comprisingof objective/short answer type questions from the entire syllabus, will be compulsory. The remaining eight questions will be set taking two questions from each unit. The candidates will be required to attempt Q.No.1 & four others selecting one question from each unit. All questions carry equal marks.

Unit	Topics	Contact Hours							
Ι	Cytopathology: General functions of the cells, cellular membranes, cytoskeleton, golgi body, lysosomes, nuclear envelope, chromatin & chromosomes, mitosis	10							
Π	Cyto-chemistry: Carbohydrates and amyloid - special stains and procedures; Connective tissues, trichrome staining and other special stains for muscle fibres, elastic, reticulin fibres and collagen fibres; Staining technique for Glycogen; Staining technique for fat; Staining technique for mucin.13								
III	Principles of metal impregnation techniques; Demonstration and identification of minerals and pigments; Metachromasia and metachromatic dyes								
IV	Cytology: Stains and cytologic preparation with special emphasis on May-Grünwald Giemsa stain MGG, and Papanicolaqu stains (PAP) method; Special stains like periodic acid Schiff (<i>PAS</i>), Mucicarmine, Alcian blue, Schmorl's, Perl's stain and Congo Red; Cytologic screening and quality control in cytology laboratory.	12							
V*	PRACTICALS 1. Study of cytological stains and staining techniques : (a) Papanicolaou staining (PAP) method (b) May-Grünwald Giemsa stain (MGG) method 2. Staining techniques for : (a) Glycogen (b) Fat (c) Mucin 3. Preparation of special cytological stains like : (a) periodic acid Schiff (PAS) (b) Mucicarmine (c) Alcian blue (d) Schmorl (e) Perl's stain (f) Congo red 4. To study staining procedures for : (a) Muscle fibres (b) Elastin fibres (c) Reticulin fibres (d) Collagen fibres 	30							

Suggested Evaluation Methods							
Internal Assessment: ➤ Theory • Class Participation: 5 • Seminar/presentation/assignment/quiz/classtestetc.:5 • Mid-Term Exam: 10 ➤ Practicum • Class Participation: NA • Seminar/Demonstration/Viva-voce/Labrecordsetc.:10 • Mid-Term Exam: NA	End Term Examination: Theory:50 (Written exam) Practical: 20 (Seminar/ Demonstration/Viva- voce/Lab records etc)						

Recommended Books/e-resources/LMS:

- 1. A Handbook of Medical Technology-second edition, By V.H. Talib, CBS Publishers
- 2. Textbook of Medical Laboratory Technology, Volume 1, 3rd Edition by Praful Ghodkar
- 3. Textbook of Medical Laboratory Technology, Volume 2, 3rd Edition by Praful Ghodkar
- 4. Medical Laboratory Technology (Volume 1): Procedure Manual for Routine Diagnostic, Kanai Mukharjee
- 5. Medical Laboratory Technology (Volume 2): Procedure Manual for routine Diagnostic, Kanai Mukharjee
- 6. Medical Laboratory Technology (Volume 3: Procedure Manual for routine Diagnostic, Kanai Mukharjee

PLOCLO Mapping of B23-MLT-403												
	PLO1	PLO2	PLO3	PLO4	PLO5	PLO6	PLO7	PLO8	PLO9			
CLO1	1	1.5	1	1	1	1.5	1	1.5	1.5			
CLO2	1.5	1	2	1	1	1.5	1	1.5	1			
CLO3	1	1.5	2	1	1	1.5	1	1.5	1.5			
CLO4	1.5	1	1	1	1	1.5	1	1.5	1			
CLO5	1	1.5	1	1	1	1.5	1	1.5	1.5			